

**INFECTIOUS DISEASE OUTBREAK LABORATORY MANUAL OF  
TESTS AND SERVICES**

**ARIZONA DEPARTMENT OF HEALTH SERVICES  
DIVISION OF PUBLIC HEALTH SERVICES  
BUREAU OF EPIDEMIOLOGY AND DISEASE CONTROL  
SERVICES  
OFFICE OF INFECTIOUS DISEASE SERVICES**

First Edition, February 10, 2011

**Overview of the Manual:**

The Arizona Department of Health Services Office of Infectious Disease Outbreak Manual of Laboratory Tests and Services contains a listing of tests performed at the Arizona State Public Health Laboratory.

The manual describes how to collect quality samples for testing and properly package them to ensure safe transport to the laboratory in the event of an outbreak.

**SECTION I** contains listings and contact numbers for epidemiology programs, specific laboratories, and emergency preparedness contacts for biological, chemical or radiological response.

**SECTION II** is a listing of reportable diseases. This list serves as a reminder to report diseases/ medical conditions to the appropriate agency within a specified time.

**SECTION III** is an alphabetized index of tests, which lists all tests performed along with the corresponding page numbers.

**SECTION IV** is an alphabetized list of testing services. Each test entry contains information pertaining to the collection, submission and analysis of samples. The entries reflect current testing capabilities and provide information for Sample, Sample and Volume, Sample Container, Sample Test Kit, Shipping Requirements and Special Instructions.

**SECTION V** contains the Arizona State Public Health Laboratory's (ASHL) Specimen Rejection policy, ASHL Microbiology Submission Form and Collection Kit request instructions

For a complete list of testing available at ASHL for routine and outbreak testing, refer to the Guide to Laboratory Services at <http://www.azdhs.gov/lab/micro/labguide.pdf>

**SECTION I- CONTACT NUMBERS FOR LABORATORY AND INFECTIOUS  
DISEASE EPIDEMIOLOGY**

**Arizona Department of Health Services  
Bureau of State Laboratory Services  
Laboratory Programs and Sections**

<b>Department</b>	<b>Phone Number</b>
<b>Receiving &amp; Stockroom</b>	<b>602 542-1190</b>
<b>Bacteriology &amp; Environmental FERN Microbiology</b>	<b>602 542-6131</b>
<b>Tuberculosis (TB)</b>	<b>602 542-6132</b>
<b>Bioemergency Detection &amp; Response</b>	<b>602 542-1150</b>
<b>Serology</b>	<b>602 542-6125 602 542-6126</b>
<b>Virology</b>	<b>602 542-6134</b>
<b>Newborn Screening</b>	<b>602 542-1184</b>
<b>Chemical Detection &amp; Response</b>	<b>602 542-3753</b>

**Arizona Department of Health Services  
Bureau of Epidemiology and Disease Control Services  
Infectious Disease Surveillance and Investigations  
Epidemiology Sections**

**IMPORTANT NOTICE:**

**The After Hours Emergency Telephone Number for the Bureau of Epidemiology  
and Disease Control Services, Office of Infectious Disease Services is  
602-839-5040**

**Call this number to report disease outbreaks, including food-borne outbreaks, cases  
of emerging infectious disease, exposures to select agents or toxins or other  
questions regarding infectious disease emergency situations that occur after hours.**

Department	Phone Number
<b>Main Number</b>	<b>602-364-3676</b>
<b>Program Manager, Surveillance Epidemiology and Investigations</b>	<b>602-364-3385</b>
<b>Foodborne/ Waterborne Epidemiologist</b>	<b>602-364-3675</b>
<b>Hepatitis B Epidemiologist</b>	<b>602-364-0246</b>
<b>Influenza Epidemiologist</b>	<b>602-319-9397</b>
<b>Investigations Epidemiologist ( Unexplained Deaths, Antibiotic Resistance, Nosocomial Infections)</b>	<b>602-364-4561</b>
<b>Laboratory Surveillance Epidemiologist</b>	<b>602-364-1442</b>
<b>Vaccine-Preventable Disease Epidemiologist</b>	<b>602-364-3685</b>
<b>Vaccine-Preventable Disease Epidemiologist</b>	<b>602-364-3817</b>

## SECTION II- LISTING OF REPORTABLE DISEASES IN ARIZONA

### COMMUNICABLE AND OTHER INFECTIOUS DISEASE REPORTABLE IN ARIZONA BY HEALTHCARE PROVIDERS

Arizona Administrative Code<sup>†</sup> Requires Providers To:  
**Report Communicable Diseases**  
 to the Local Health Department

<input type="checkbox"/> * <input type="radio"/> Amebiasis	<input type="checkbox"/> Hantavirus infection	<input type="checkbox"/> * <input type="radio"/> Salmonellosis
<input type="checkbox"/> Anthrax	<input type="checkbox"/> Hemolytic uremic syndrome	<input type="radio"/> Scabies
<input type="checkbox"/> Aseptic meningitis: viral	<input type="checkbox"/> * <input type="radio"/> Hepatitis A	<input type="checkbox"/> Severe acute respiratory syndrome
<input type="checkbox"/> Basidiobolomycosis	<input type="checkbox"/> Hepatitis B and D	<input type="checkbox"/> * <input type="radio"/> Shigellosis
<input type="checkbox"/> Botulism	<input type="checkbox"/> Hepatitis C	<input type="checkbox"/> Smallpox
<input type="radio"/> Brucellosis	<input type="checkbox"/> * <input type="radio"/> Hepatitis E	<input type="checkbox"/> Streptococcal Group A: invasive disease
<input type="checkbox"/> * <input type="radio"/> Campylobacteriosis	<input type="checkbox"/> Herpes genitalis	<input type="checkbox"/> Streptococcal Group B: invasive disease in infants younger than 90 days of age
<input type="checkbox"/> Chagas disease (American trypanosomiasis)	<input type="checkbox"/> HIV infection and related disease	<input type="checkbox"/> <i>Streptococcus pneumoniae</i> (pneumococcal invasive disease)
<input type="checkbox"/> Chancre	<input type="radio"/> Influenza-associated mortality in a child	<input type="checkbox"/> Syphilis
<input type="checkbox"/> Chlamydia infection, sexually transmitted	<input type="checkbox"/> Kawasaki syndrome	<input type="checkbox"/> * <input type="radio"/> Taeniasis
<input type="radio"/> * Cholera	<input type="checkbox"/> Legionellosis (Legionnaires' disease)	<input type="checkbox"/> Tetanus
<input type="checkbox"/> Coccidioidomycosis (valley fever)	<input type="checkbox"/> Leptospirosis	<input type="checkbox"/> Toxic shock syndrome
<input type="checkbox"/> Colorado tick fever	<input type="checkbox"/> Listeriosis	<input type="checkbox"/> Trichinosis
<input type="radio"/> Conjunctivitis: acute	<input type="checkbox"/> Lyme disease	<input type="radio"/> Tuberculosis, active disease
<input type="checkbox"/> Creutzfeldt-Jakob disease	<input type="checkbox"/> Lymphocytic choriomeningitis	<input type="radio"/> Tuberculosis latent infection in a child 5 years of age or younger (positive screening test result)
<input type="checkbox"/> * <input type="radio"/> Cryptosporidiosis	<input type="checkbox"/> Malaria	<input type="checkbox"/> Tularemia
<input type="checkbox"/> <i>Cyclospora</i> infection	<input type="checkbox"/> Measles (rubeola)	<input type="checkbox"/> Typhoid fever
<input type="checkbox"/> Cysticercosis	<input type="checkbox"/> Meningococcal invasive disease	<input type="checkbox"/> Typhus fever
<input type="checkbox"/> Dengue	<input type="radio"/> Mumps	<input type="checkbox"/> Unexplained death with a history of fever
<input type="radio"/> Diarrhea, nausea, or vomiting	<input type="checkbox"/> Pertussis (whooping cough)	<input type="radio"/> Vaccinia-related adverse event
<input type="checkbox"/> Diphtheria	<input type="checkbox"/> Plague	<input type="checkbox"/> Vancomycin-resistant or Vancomycin-intermediate <i>Staphylococcus aureus</i>
<input type="checkbox"/> Ehrlichiosis and Anaplasmosis	<input type="checkbox"/> Poliomyelitis	<input type="checkbox"/> Vancomycin-resistant <i>Staphylococcus epidermidis</i>
<input type="checkbox"/> Emerging or exotic disease	<input type="checkbox"/> Psittacosis (ornithosis)	<input type="checkbox"/> Varicella (chickenpox)
<input type="radio"/> Encephalitis, viral or parasitic	<input type="radio"/> Q fever	<input type="checkbox"/> * <input type="radio"/> <i>Vibrio</i> infection
<input type="checkbox"/> Enterohemorrhagic <i>Escherichia coli</i>	<input type="checkbox"/> Rabies in a human	<input type="checkbox"/> Viral hemorrhagic fever
<input type="checkbox"/> Enterotoxigenic <i>Escherichia coli</i>	<input type="checkbox"/> Relapsing fever (borreliosis)	<input type="checkbox"/> West Nile virus infection
<input type="checkbox"/> * <input type="radio"/> Giardiasis	<input type="checkbox"/> Reye syndrome	<input type="checkbox"/> Yellow fever
<input type="checkbox"/> Gonorrhea	<input type="checkbox"/> Rocky Mountain spotted fever	<input type="checkbox"/> * <input type="radio"/> Yersiniosis
<input type="checkbox"/> <i>Haemophilus influenzae</i> : invasive disease	<input type="radio"/> * Rubella (German measles)	
<input type="checkbox"/> Hansen's disease (Leprosy)	<input type="radio"/> Rubella syndrome, congenital	

Submit a report by telephone or through an electronic reporting system authorized by the Department within 24 hours after a case or suspect case is diagnosed, treated, or detected or an occurrence is detected.

\* If a case or suspect case is a food handler or works in a child care establishment or a health care institution, instead of reporting within the general reporting deadline, submit a report within 24 hours after the case or suspect case is diagnosed, treated, or detected.

Submit a report within one working day after a case or suspect case is diagnosed, treated, or detected.

Submit a report within five working days after a case or suspect case is diagnosed, treated, or detected.

Submit a report within 24 hours after detecting an outbreak.

## COMMUNICABLE AND OTHER INFECTIOUS DISEASE REPORTABLE IN ARIZONA BY ALL CLINICAL LABORATORIES

### Reports should be sent to:

Arizona Department of Health Services  
 Infectious Disease Epidemiology  
 150 North 18<sup>th</sup> Avenue, Suite 140  
 Phoenix, AZ 85007  
 602-364-3676 or 602-364-3199 (fax)

## ARIZONA LABORATORY REPORTING REQUIREMENTS

### Isolates should be sent to:

Arizona State Laboratory  
 250 North 17<sup>th</sup> Avenue  
 Phoenix, AZ 85007

①	Arboviruses	☒*	<i>Haemophilus influenzae</i> , other, isolated from a normally sterile site	☒	<i>Plasmodium</i> spp.
☒☒*	<i>Bacillus anthracis</i>	☒	Hantavirus	☒	Respiratory syncytial virus
☒*	<i>Bordetella pertussis</i>	☒ <sup>1</sup>	Hepatitis A virus (anti-HAV-IgM serologies)	☒+	Rubella virus and anti-rubella-IgM serologies
☒*	<i>Brucella</i> spp.	☒ <sup>1</sup>	Hepatitis B virus (anti-Hepatitis B core-IgM serologies, Hepatitis B surface or envelope antigen serologies, or detection of viral nucleic acid)	☒*	<i>Salmonella</i> spp.
☒*	<i>Burkholderia mallei</i> and <i>B. pseudomallei</i>	☒ <sup>1</sup>	Hepatitis C virus	☒	SARS-associated corona virus
☒	<i>Campylobacter</i> spp.	☒ <sup>1</sup>	Hepatitis D virus	☒*	<i>Shigella</i> spp.
☒	CD <sub>4</sub> -T-lymphocyte count of fewer than 200 per microliter of whole blood or CD <sub>4</sub> -T-lymphocyte percentage of total lymphocytes of less than 14%	☒ <sup>1</sup> +	Hepatitis E virus (anti-HEV-IgM serologies)	☒	<i>Streptococcus</i> Group A, isolated from a normally sterile site
☒	<i>Chlamydia trachomatis</i>	☒	HIV (by culture, antigen, antibodies to the virus, or detection of viral nucleic acid)	☒	<i>Streptococcus</i> Group B, isolated from a normally sterile site in an infant younger than 90 days of age
☒☒	<i>Clostridium botulinum</i> toxin (botulism)	☒	HIV—any test result for an infant (by culture, antigen, antibodies to the virus, or detection of viral nucleic acid)	☒*	<i>Streptococcus pneumoniae</i> and its drug sensitivity pattern, isolated from a normally sterile site
☒	<i>Coccidioides</i> spp., by culture or serologies	☒	Influenza virus	☒	<i>Treponema pallidum</i> (syphilis)
☒	<i>Coxiella burnetii</i>	☒*	<i>Legionella</i> spp. (culture or DFA)	☒	<i>Trypanosoma cruzi</i> (Chagas disease)
☒	<i>Cryptosporidium</i> spp.	☒*	<i>Listeria</i> spp., isolated from a normally sterile site	☒*	Vancomycin-resistant or Vancomycin-intermediate <i>Staphylococcus aureus</i>
☒	<i>Cyclospora</i> spp.	☒+	Measles virus and anti-measles-IgM serologies	☒*	Vancomycin resistant <i>Staphylococcus epidermidis</i>
☒	Dengue virus	☒ <sup>2</sup>	Methicillin-resistant <i>Staphylococcus aureus</i> , isolated from a normally sterile site	☒☒	Variola virus (smallpox)
☒☒	Emerging or exotic disease agent	☒+	Mumps virus and anti-mumps-IgM serologies	☒*	<i>Vibrio</i> spp.
☒	<i>Entamoeba histolytica</i>	☒*	<i>Mycobacterium tuberculosis</i> complex and its drug sensitivity pattern	☒☒	Viral hemorrhagic fever agent
☒	<i>Escherichia coli</i> O157:H7	☒	<i>Neisseria gonorrhoeae</i>	☒	West Nile virus
☒*	<i>Escherichia coli</i> , Shiga-toxin producing	☒*	<i>Neisseria meningitidis</i> , isolated from a normally sterile site	☒*	<i>Yersinia</i> spp. (other than <i>Y. pestis</i> )
☒☒*	<i>Francisella tularensis</i>	☒	Norovirus	☒☒*	<i>Yersinia pestis</i> (plague)
☒*	<i>Haemophilus influenzae</i> , type b, isolated from a normally sterile site				

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- ☒ Submit a report immediately after receiving one specimen for detection of the agent. Report receipt of subsequent specimens within five working days after receipt.
  - ☒ Submit a report within 24 hours after obtaining a positive test result.
  - ① Submit a report within one working day after obtaining a positive test result.
  - ☒ Submit a report within five working days after obtaining a positive test result or a test result specified on this page.
  - \* Submit an isolate of the organism for each positive culture to the Arizona State Laboratory at least once each week, as applicable.
  - + For each positive test result, submit a specimen to the Arizona State Laboratory within 24 hours after obtaining the positive test result.
  - <sup>1</sup> When reporting a positive result for any of the specified tests, report the results of all other tests performed for the subject as part of the disease panel.
  - <sup>2</sup> Submit a report only when an initial positive result is obtained for an individual.
  - <sup>3</sup> Submit an isolate of the organism only when an initial positive result is obtained for an individual, when a change in resistance pattern is detected, or when a positive result is obtained ≥ 12 months after the initial positive result is obtained for an individual.

[www.azdhs.gov/phs/oids/lab\\_rpt.htm](http://www.azdhs.gov/phs/oids/lab_rpt.htm)

**A.A.C. R9-6-204**  
 Effective 04/01/2008

### SECTION III: INDEX OF TESTS FOR INFECTIOUS DISEASE COMMONLY ASSOCIATED WITH OUTBREAKS

TEST CATEGORY	TEST NAME	PAGE
<i>Bordetella pertussis</i>	<ul style="list-style-type: none"> <li>▪ <i>Bordetella pertussis</i> Culture</li> <li>▪</li> </ul>	<b>8</b>
<b>Botulism</b>	<ul style="list-style-type: none"> <li>▪ Botulism Toxin Culture, Food or Stool</li> </ul>	<b>9</b>
<b>Campylobacter</b>	<ul style="list-style-type: none"> <li>▪ <i>Campylobacter</i> spp. Culture</li> <li>▪ <i>Campylobacter</i> spp. Isolation, Food</li> </ul>	<b>10</b>
<b>Cryptosporidium</b>	<ul style="list-style-type: none"> <li>▪ Intestinal Parasite Exam</li> </ul>	<b>11</b>
<b>Enteric Culture</b>	<ul style="list-style-type: none"> <li>▪ General Enteric Culture- Isolation and identification of <i>Shigella</i> spp., <i>Salmonella</i> spp., <i>Camoylobacter</i> spp., and toxin producing <i>E.coli</i>.</li> </ul>	
<b>Escherichia coli</b>	<ul style="list-style-type: none"> <li>▪ Enterohemorrhagic <i>E.coli</i> (EHEC) O157:H7 Culture</li> <li>▪ Enterohemorrhagic <i>E.coli</i> (EHEC) NON-O157:H7 Serogrouping</li> <li>▪ Shiga Toxin-Producing <i>E.coli</i> (STEC) PCR</li> </ul>	<b>12</b>
<b>Giardia</b>	<ul style="list-style-type: none"> <li>▪ Intestinal Parasite Exam (O &amp; P)</li> </ul>	<b>13</b>
<i>Haemophilus influenzae</i>	<ul style="list-style-type: none"> <li>▪ <i>Haemophilus influenzae</i> Biotyping &amp; Serotyping</li> </ul>	<b>14</b>
<b>Hepatitis A</b>	<ul style="list-style-type: none"> <li>▪ Hepatitis A, IgM EIA</li> </ul>	<b>15</b>
<b>Influenza</b>	<ul style="list-style-type: none"> <li>▪ Influenza Culture</li> <li>▪ Influenza PCR</li> </ul>	<b>16</b>
<b>Legionella</b>	<ul style="list-style-type: none"> <li>▪ <i>Legionella</i> spp. Culture</li> <li>▪ <i>Legionella</i> spp. Serogrouping</li> </ul>	<b>17</b>
<b>Listeria</b>	<ul style="list-style-type: none"> <li>▪ <i>Listeria</i> spp. Culture</li> <li>▪ <i>Listeria monocytogenes</i> Isolation, Food</li> </ul>	<b>18</b>
<b>Measles</b>	<ul style="list-style-type: none"> <li>▪ Measles, IgM EIA</li> <li>▪ Measles PCR</li> </ul>	<b>19-20</b>
<b>Mumps</b>	<ul style="list-style-type: none"> <li>▪ Mumps, IgM EIA</li> <li>▪ Mumps PCR</li> </ul>	<b>21-22</b>
<i>Neisseria meningitidis</i>	<ul style="list-style-type: none"> <li>▪ <i>Neisseria meningitidis</i> Culture &amp; Serogrouping</li> </ul>	<b>23</b>
<b>Norovirus</b>	<ul style="list-style-type: none"> <li>▪ Norovirus PCR</li> </ul>	<b>24</b>
<b>Rubella</b>	<ul style="list-style-type: none"> <li>▪ Rubella IgM EIA</li> </ul>	<b>25-26</b>
<b>Salmonella</b>	<ul style="list-style-type: none"> <li>▪ <i>Salmonella</i> spp. Culture</li> <li>▪ <i>Salmonella</i> spp. Isolation, Food</li> </ul>	<b>27</b>
<b>Shigella</b>	<ul style="list-style-type: none"> <li>▪ <i>Shigella</i> spp. Culture</li> <li>▪ <i>Shigella</i> spp. Isolation, Food</li> </ul>	<b>28</b>
<b>Vibrio</b>	<ul style="list-style-type: none"> <li>▪ <i>Vibrio</i> spp. Culture</li> </ul>	<b>29</b>
<b>Bacterial Typing</b>	<ul style="list-style-type: none"> <li>▪ Bacterial Typing, Pulsed Field Gel Electrophoresis (PFGE)</li> </ul>	<b>30</b>

## SECTION IV: LABORATORY TESTING SERVICES

### *Bordetella pertussis*

<b>Test Name:</b>	<b><u><i>Bordetella pertussis</i> Culture</u></b>
<b>Use of Test:</b>	Isolation and identification of <i>Bordetella pertussis</i> .
<b>Significant Result:</b>	Culture positive for <i>Bordetella pertussis</i> .
<b>Limitations:</b>	Culture is most sensitive for specimens collected within the first 3 weeks after cough onset. Beyond this point, false negative results become more common.
<b>Turnaround Time:</b>	11 days
<b>Sample and Volume:</b>	Nasopharyngeal swab
<b>Sample Collection:</b>	<b><u>Nasopharyngeal swab:</u></b> The pharynx is swabbed vigorously with a swab. <b><u>USE ONLY DACRON SWABS.</u></b> Specimens submitted should not be frozen. Post collection, push the swab into a tube of Regan-Lowe semi-solid transport agar or Amies with charcoal.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	The specimens should be shipped on ice. If transport to ASHL is delayed, specimens should be refrigerated. Use a triple packaging system for transporting by courier or USPS. Mark the outer packaging “UN3373- Biological Substances, Category B.”

## **Botulism**

**Test Name:**

**Special Instructions:**

**Use of Test:**

**Turnaround Time:**

**Sample and Volume:**

**Botulism Toxin Culture, Food or Stool**

ALL BOTULISM TESTING IS REFERRED TO ADHS, BUREAU OF EPIDEMIOLOGY AND DISEASE CONTROL PRIOR TO SENDING SPECIMENS IN ORDER TO ALERT STAFF. IF AFTER HOURS PLEASE CONTACT THE EPIDEMIOLOGY 24/7 ON-CALL NUMBER AT **602-920-3772** AND THE STATE LABORATORY 24/7 ON-CALL NUMBER AT **602-283-6277**

To support a diagnosis of botulism, infant botulism.

Test performed at CDC

**Infant Botulism:**

1. Stool for culture and toxin- 20 to 50 grams
2. Serum for toxin- 2.5 ml minimum
  - Toxin testing- 10 to 30 grams
  - Culture- 10 to 20 grams or 15 to 25ml of watery enema.
  - A rectal swab may be accepted, only if other stool specimens are not available.
3. Food for toxin and culture

**Food borne Botulism- Adult:**

1. Feces- 25 to 50 grams
2. Serum – 15 to 20 ml
3. Remainder of suspected food

**Wound Botulism:**

1. Feces- 25 to 50 grams
2. Serum – 15 to 20 ml
3. Tissue, exudates or swab samples from wound

**Sample Container:**

**Forms Required:**

**Shipping Requirements:**

Sterile, leak proof container and insulated box with coolant. **DO NOT FREEZE.**

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

Shipment by courier as soon as possible is optimal. If necessary, ship overnight with coolant. **DO NOT FREEZE.** Triple package sample as an “UN3373-Biological Substances, Category B” for transport by Courier or USPS.

## Campylobacter

<b>Test Name:</b>	<b><u>Campylobacter spp. Culture</u></b>
<b>Use of Test:</b>	To screen for bacterial cause of diarrheal illness
<b>Special Instructions:</b>	Stool specimens must be properly submitted, with transport containers not overfilled and with transport medium not removed. Specimen jars must be tightly closed and not leaking when received. The time interval between collection of the specimen and receipt in the Lab must not be greater than 5 days.
<b>Test Includes:</b>	Isolation and species identification of <i>Campylobacter</i> species. All <i>Campylobacter</i> isolates are sent to CDC for serotyping. Tests also include Pulsed-field gel electrophoresis (PFGE).
<b>Turnaround Time:</b>	5 days
<b>Sample and Volume:</b>	Stool specimen (approximately one gram). Stool swab and rectal swab are acceptable but less desirable than stool.
<b>Sample Collection:</b>	<i>Isolate-</i> In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture on a suitable nutrient agar slant. TSI is <b>not</b> to be used for <i>Campylobacter</i> spp. <i>Specimen:</i> Collect a small portion of feces, approximately the size of a marble, or a swab coated with feces and place in Cary-Blair transport media. Inoculate a small amount of stool below the surface of the medium.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type. The State Laboratory will provide agencies with Cary-Blair Medium.
<b>Shipping Requirements:</b>	Specimens held in Cary-Blair Medium should be refrigerated until examined. Use triple packaging system for courier or USPS. Mark outer packaging as “UN3373-Biological Substances, Category B”.

<b>Test Name:</b>	<b><u>Campylobacter spp. Isolation, Food</u></b>
<b>Use of Test:</b>	To support epidemiologic evidence implicating a food as a possible source of illness.
<b>Special Instructions:</b>	Food samples must be submitted through local or state public health agencies as part of an outbreak investigation (one or more ill consumers). All food items must clearly be labeled with the name of the food source and date of collection
<b>Test Includes:</b>	Isolation and identification of <i>Campylobacter</i> species
<b>Turnaround Time:</b>	3 to 7 days.
<b>Sample and Volume:</b>	At least 200 grams of the solid product or 100 ml of liquid.
<b>Sample Container:</b>	Original sample container as submitted by inspector or other sterile leak proof container.
<b>Sample Collection:</b>	Collect food aseptically and place in sterile whirlpack bags or other sterile, leak proof container. Keep all samples refrigerated except those samples received frozen which should be maintained in the frozen state.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Triple package samples. Transport samples on ice or on prefrozen cold packs, in appropriate packaging.

## *Cryptosporidium*

**Test Name:**

**Use of Test:**

**Special Instructions:**

**Test Includes:**

**Turnaround Time:**

**Sample and Volume:**

**Sample Collection:**

### *Cryptosporidiosis spp. Detection*

Detection of *Cryptosporidium* oocysts in stool.

Stool specimens must be properly submitted, with transport containers not overfilled and with transport medium not removed. Specimen jars must be tightly closed and not leaking when received. Because the host passes parasites intermittently, multiple specimens should be examined. These irregularities emphasize the need to collect at least three specimens over 10-14 days.

The detection of 1 or more *Cryptosporidium* oocysts microscopically.

5 days

Stool specimen (approximately one gram).

*Specimen:* Collect the stool in a clean container or on a clean paper and transfer to the Ova and Parasite transport containers supplied by the State Laboratory. The collection kit provided includes a container with PVA fixative and one container with 10% formalin fixative. A portion of the specimen, approximately 1 tablespoon, is added to the fixative in a ratio of 1 part specimen to 3 parts fixative. Mix thoroughly to assure adequate fixation. Do not contaminate specimen with dirt or urine.

**TREATMENT WITH ANY OF THE FOLLOWING BEFORE COLLECTION WILL MAKE SPECIMENS UNSUITABLE FOR TESTING: ANTACIDS, BISMUTH, ANTI-DIARRHEAL MEDICATIONS, ANTIBIOTICS, AND/OR OILY LAXATIVES.**

**Forms Required:**

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

**Shipping Requirements:**

Mailed stool specimens require use of a preservative, and a two-vial method of collection. One vial should contain 10% formalin and the other contains PVA fixative. Use triple packaging system for courier or USPS. Mark outer packaging as "UN3373-Biological Substances, Category B".

## Enteric culture

**Test Name:**

**Use of Test:**

**Test includes:**

**Special Instructions:**

**Turnaround Time:**

**Sample and Volume:**

**Sample Collection:**

**Forms Required:**

**Shipping Requirements:**

**Enteric culture**

To screen for bacterial cause of diarrheal illness

Isolation and identification of *Shigella* spp., *Salmonella* spp., *Campylobacter* spp., and toxin producing *E.coli*. If isolate is identified and problematic then isolates are submitted to CDC for serotyping. Tests also include pulsed-field gel electrophoresis (PFGE).

Stool specimens must be properly submitted, with transport containers not overfilled and with transport medium not removed. Specimen jars must be tightly closed and not leaking when received. The time interval between collection of the specimen and receipt in the Lab must not be greater than 5 days.

10 days

Stool specimen (approximately one gram)

*Specimen*- If isolate not available, then collection of stool in Cary-Blair transport media is required for testing.

For Cary-Blair Medium, inoculate a small amount of stool below the surface of the medium. The ASHL will provide agencies with Cary-Blair Medium.

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

Specimens held in Cary-Blair Medium should be refrigerated until examined. Samples older than 3 days will not be accepted. Use triple packaging system for courier or USPS. Mark outer packaging as "UN3373-Biological Substances, Category B".

## Escherichia coli (E.coli)

<b>Test Name:</b>	<b><u>Enterohemorrhagic E.coli (EHEC) O157:H7 Culture</u></b>
<b>Use of Test:</b>	To screen for bacterial cause of diarrheal illness
<b>Test includes:</b>	Isolation and identification of <u>Enterohemorrhagic E.coli</u> . Tests include identification of <u>E.coli</u> O157:H7 serogroup. Problematic isolates are submitted to CDC for serotyping. Tests also include pulsed-field gel electrophoresis (PFGE).
<b>Special Instructions:</b>	Stool specimens must be properly submitted, with transport containers not overfilled and with transport medium not removed. Specimen jars must be tightly closed and not leaking when received. The time interval between collection of the specimen and receipt in the Lab must not be greater than 5 days.
<b>Turnaround Time:</b>	5 days
<b>Sample and Volume:</b>	Stool specimen (approximately one gram)
<b>Sample Collection:</b>	<i>Isolate or Broth-</i> In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture on TSI or nutrient agar slant or culture plate <i>Specimen-</i> If isolate not available, then collection of stool in Cary-Blair transport media is required for testing. For Cary-Blair Medium, inoculate a small amount of stool below the surface of the medium. The ASHL will provide agencies with Cary-Blair Medium.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Specimens held in Cary-Blair Medium should be refrigerated until examined. Samples older than 3 days will not be accepted. Use triple packaging system for courier or USPS. Mark outer packaging as “UN3373-Biological Substances, Category B”.
<b>Test Name:</b>	<b><u>Enterohemorrhagic E.coli (EHEC) NON-O157:H7 Serogrouping</u></b> <b><u>ShigaToxin-Producing E.coli (STEC) PCR</u></b>
<b>Use of Test:</b>	Confirm presence of Shiga toxin. Isolate Shiga-toxin producing organism(s) for subsequent identification.
<b>Test Includes:</b>	Test for Shiga toxin(s) by Polymerase chain reaction (PCR). Isolation of Shiga-toxin producing organism from positive specimens for subsequent identification. Confirmation of suspected Shiga toxin-producing <i>E.coli</i> (STEC) or other suspected Shiga toxin producing organism and subsequent serotyping.
<b>Turnaround Time:</b>	5-7 days for confirmation of culture and/or stool specimen. Successful isolation of the Shiga-toxin producing organism can take a few days longer. For final confirmation and serotyping, the isolates may be sent to CDC. Results maybe over a month from time specimen submitted to CDC.
<b>Sample and Volume:</b>	Stool specimen (5-25 grams)
<b>Sample Collection:</b>	<i>Isolate or Broth-</i> In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture, collect on an agar slant or plate. <i>Specimen-</i> If isolate not available, then collection of stool in Cary-Blair transport media is required for testing. For Cary-Blair Medium, inoculate a small amount of stool below the surface of the medium. The ASHL will provide agencies with Cary-Blair Medium.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Ship pure cultures at room temperature. Specimens held in Cary-Blair Medium should be refrigerated until examined. Use triple packaging system for courier or USPS. Mark outer packaging as “UN3373-Biological Substances, Category B”.

## Giardia

**Test Name:**

### *Giardiasis spp. Detection*

**Use of Test:**

Detection of Giardia oocysts in stool.

**Special Instructions:**

Stool specimens must be properly submitted, with transport containers not overfilled and with transport medium not removed. Specimen jars must be tightly closed and not leaking when received. It is important to collect at least three specimens over 10-14 days.

**Test Includes:**

The detection of 1 or more Giardia oocysts microscopically.

**Turnaround Time:**

5 days

**Sample and Volume:**

Stool specimen (approximately one gram).

**Sample Collection:**

*Specimen:* Collect the stool in a clean container or on a clean paper and transfer to the Ova and Parasite transport containers supplied by the State Laboratory. The collection kit provided includes a container with PVA fixative and one container with 10% formalin fixative. A portion of the specimen, approximately 1 tablespoon, is added to the fixative in a ratio of 1 part specimen to 3 parts fixative. Mix thoroughly to assure adequate fixation. Do not contaminate specimen with dirt or urine.

**TREATMENT WITH ANY OF THE FOLLOWING BEFORE COLLECTION WILL MAKE SPECIMENS UNSUITABLE FOR TESTING: ANTACIDS, BISMUTH, ANTI-DIARRHEAL MEDICATIONS, ANTIBIOTICS, AND/OR OILY LAXATIVES.**

**Forms Required:**

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type. Mailed stool specimens require use of a preservative, and a two-vial method of collection. One vial should contain 10% formalin and the other contains PVA fixative. Use triple packaging system for courier or USPS. Mark outer packaging as “UN3373-Biological Substances, Category B”.

**Shipping Requirements:**

## *Haemophilus influenzae*

**Test Name:**

### *Haemophilus influenzae* Biotyping & Serotyping

**Test Includes:**

Biotyping and serotyping of *Haemophilus influenzae*

**Limitations:**

Testing performed only on organisms isolated from normally sterile sites.

**Turnaround Time:**

5 days

**Sample:**

Pure culture on chocolate agar slant or culture plate.

**Specimen Collection:**

*Isolate-* In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture on suitable chocolate agar slant or culture plate.

### Forms Required:

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

**Shipment Requirements:**

Ship at room temperature. Using the triple packaging system for courier or USPS, mark outer packaging as “UN3373-Biological Substances, Category B.”

## **Hepatitis A**

**Test Name:**

**Hepatitis A IgM (EIA)**

**Use of Test:**

Detection of antibody to Hepatitis A IgM antibody

**Test Includes:**

Qualitative testing by a commercial enzyme immunoassay (EIA) procedure.

**Turnaround Time:**

4 days

**Sample Collection:**

Routine blood draw. Minimum of 1mL serum or plasma needed. Serum separator tube preferred, though red top serum tubes are acceptable.

**Forms Required:**

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

**Shipping Requirements:**

Use triple packaging system for courier or USPS. Mark outer packaging as “UN3373-Biological Substances, Category B”.

## **Influenza**

**Test Name:**

**Test Includes:**

**Turnaround Time:**

**Specimen:**

### **Influenza Culture & Typing/Subtyping**

Isolation and typing of influenza virus by shell vials.

14 days

*Preferred*- Nasopharyngeal swab

*Acceptable*- Nasopharyngeal wash or aspirate, throat swab, tracheal aspirate, sputum, bronchoalveolar lavage (BAL).

**Specimen Collection:**

Virus isolation is most successful if specimen collected within 3 to 5 days of illness. The pharynx is swabbed vigorously with a cotton swab moistened with collection medium free of serum such as Hanks, and then placed in a transport container containing Hanks Buffered Saline Solution (HBSS). Specimens submitted should not be frozen.

**Forms Required:**

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

**Shipping Requirements:**

Shipment by courier or transport to the ASHL within 24 hours on prefrozen ice packs to keep refrigerated. Use a triple packaging system for transporting by courier. Mark the outer packaging as "UN3373-Biological Substances, Category B."

**Test Name:**

**Test Includes:**

**Turnaround Time:**

**Specimen:**

### **Influenza, Molecular Typing PCR**

Identification of influenza types A, B and subtypes H1, H3, H5, and 2009 H1N1 by Polymerase Chain Reaction (PCR).

2 days

*Preferred*- Nasopharyngeal swab

*Acceptable*- Nasopharyngeal wash or aspirate, throat swab, tracheal aspirate, sputum, bronchoalveolar lavage (BAL).

**Specimen Collection:**

Virus isolation is most successful if specimen collected within 3 to 5 days of illness.

Nasopharyngeal swab: The pharynx is swabbed vigorously with a cotton swab moistened with collection medium free of serum such as Hanks, and then placed in a transport container containing Hanks Buffered Saline Solution (HBSS). Specimens submitted should not be frozen.

**Forms Required:**

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

**Shipping Requirements:**

Shipment by courier or transport to the ASHL within 24 hours on prefrozen ice packs to keep refrigerated. Use a triple packaging system for transporting by courier. Mark the outer packaging as "UN3373-Biological Substances, Category B."

## **Legionellosis (Legionnaires' Disease)**

<b>Test Name:</b>	<b><u>Legionella Culture &amp; Serogrouping</u></b>
<b>Use of Test:</b>	Identification and serogrouping of <i>Legionella</i> spp.
<b>Turnaround Time:</b>	7 days for species identification. Sent to CDC for serotyping
<b>Specimen:</b>	Pleural fluid, transtracheal aspirate, respiratory secretions (sputum, bronchial wash, bronchoalveolar lavage (BAL), and normally sterile sites.
<b>Specimen Collection:</b>	<i>Isolate</i> -In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture on BCYE agar plate <i>Specimen</i> -Collect 5 to 30mL of secretions. Specimens should be held at 4-8°C and should not be allowed to dry out. DO NOT use sterile saline for specimen collections as <i>Legionella</i> spp. are inhibited by saline.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	The specimens should be shipped on ice the same day. If transport to ASHL is delayed, specimens should be frozen (-70°C) and transported in the frozen state (dry ice). Use a triple packaging system for transporting by courier or USPS. Mark the outer packaging “UN3373- Biological Substances, Category B.”

## Listeria

**Test Name:**

**Use of Test:**

**Special Instructions:**

**Test Includes:**

**Turnaround Time:**

**Sample Collection:**

**Forms Required:**

**Shipping Requirements:**

**Test Name:**

**Use of Test:**

**Special Instructions:**

**Test Includes:**

**Turnaround Time:**

**Sample and Volume:**

**Sample Container:**

**Sample Collection:**

**Forms Required:**

**Shipping Requirements:**

**Listeria spp. Culture**

Identification of *Listeria* spp.

Clinical specimens from normally sterile sites such as blood, cerebrospinal fluid (CSF), amniotic fluid, placenta, or fetal tissue do not require special procedures for collection or transport. Specimens from non-sterile sites, such as meconium, feces, vaginal secretions, respiratory, skin or mucous swabs require prompt handling to prevent overgrowth of contaminants.

Confirmation of isolates. Tests also include pulsed-field gel electrophoresis (PFGE). All isolates of *Listeria* spp. are sent to CDC for serotyping

5 days for species identification. Sent to CDC for serotyping

*Isolate-* In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture from a sterile site that can be plated directly on tryptic soy agar containing 5% sheep, horse, or rabbit blood. Samples for blood culture can be inoculated directly into conventional blood culture broth.

*Specimen-* If isolates not available, then collection of Blood or CSF is required for testing. No transport media is required, but specimens should be shipped at 4°C

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

Specimens from sterile sites should be shipped as soon as possible.

If processing is delayed, specimens should be held at 35°C in an incubator for no longer than 48 hours. Ship at 4°C. Specimens from non-sterile sites require prompt handling. If processing is delayed, the materials should be kept at 4°C or frozen at -20°C if testing delays are to exceed 48 hours. Ship at 4°C.

Stools should be shipped frozen on dry ice.

Use triple packaging system for courier or USPS. Mark outer packaging as "UN3373-Biological Substances, Category B".

**Listeria monocytogenes Isolation, Food**

To support epidemiologic evidence implicating a food as a possible source of illness.

Food samples must be submitted through local or state public health agencies as part of an outbreak investigation (one or more ill consumers). Epidemiology needs to be notified of any food samples being sent for testing. ASL has limited media available on hand and needs to be notified by Epidemiology. All food items must clearly be labeled with the name of the food source and date of collection

Isolation and identification of *Listeria monocytogenes*.

3 to 7 days.

At least 200 grams of the solid product or 100 ml of liquid.

Original sample container as submitted by inspector or other sterile leak proof container. Collect food aseptically and place in sterile whirlpack bags or other sterile, leak proof container. Keep all samples refrigerated except those samples received frozen which should be maintained in the frozen state.

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

Triple package samples. Transport samples on ice or on pre-frozen cold packs, in appropriate packaging.

## Measles

<b>Test Name:</b>	<b><u>Measles Antibody IgM (EIA)</u></b>
<b>Use of Test:</b>	Determination of measles infection.
<b>Significant Result:</b>	Positive IgM indicates current or recent measles infection.
<b>Limitations:</b>	IgM may be negative if the specimen is collected prior to the appearance of or before the third day after rash onset. Cannot distinguish between antibody produced in response to vaccine versus wild strain measles.
<b>Turnaround Time:</b>	3 days
<b>Sample and Volume:</b>	Whole blood (10-15mL) collected aseptically in a red top vacutainer tube or single serum (2-3mL).
<b>Sample Collection:</b>	Serum collected 3-7 days after the appearance of rash.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Due to the intense heat in the summertime, it is advisable to ship the specimens cold to prevent damage to the specimen. Whole blood samples may be sent on cool packs and not frozen. Serum samples, if not tested within 7 days or already frozen, should be shipped frozen to ASHL on dry ice. <u>If sent by courier:</u> <ol style="list-style-type: none"><li>1. Blood and blood products sent in vacutainer tubes should first be placed in a plastic falcon tube.</li><li>2. The specimen should then be placed in a plastic specimen bag with separate compartments for the submission form and the specimen.</li><li>3. Pack the specimen and its form in absorbent material</li><li>4. Mark the outer packaging “UN3373- Biological Substances, Category B.”</li></ol> <u>If sent by mail:</u> <ol style="list-style-type: none"><li>1. Blood and blood products sent in vacutainer tubes should first be placed in a plastic falcon tube.</li><li>2. Check with the Post Office for current postage requirements.</li><li>3. Wrap the submission form around the falcon tube, and place the falcon tube inside a Styrofoam container or cardboard mailer. Pack the specimen in absorbent material.</li><li>4. Mark the outer packaging “UN3373- Biological Substances, Category B.”</li></ol>

<b>Test Name:</b>	<b><u>Measles, Molecular Typing PCR</u></b>
<b>Use of Test:</b>	Detection of measles RNA virus
<b>Turnaround Time:</b>	2 days
<b>Sample:</b>	Nasopharyngeal swab, throat swab, whole blood, and urine (5mL).
<b>Sample Collection:</b>	<u>Nasopharyngeal swab:</u> The pharynx is swabbed vigorously with a cotton swab moistened with collection medium free of serum such as Hanks, and then placed in a transport container containing Hanks Buffered Saline Solution (HBSS). Specimens submitted should not be frozen. <u>Throat Swab:</u> The pharynx is swabbed vigorously with a cotton swab moistened with collection medium free of serum such as Hanks, and then placed in a transport container containing Hanks Buffered Saline Solution (HBSS). Samples submitted should not be frozen. <u>Urine:</u> Clean catch urine collected in a sterile container.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Use a triple packaging system for transporting by courier. Mark the outer packaging as “UN3373- Biological Substances, Category B.” <b><u>Ship cold</u></b>

## Mumps

<b>Test Name:</b>	<b><u>Mumps Antibody IgM (EIA)</u></b>
<b>Use of Test:</b>	Determination of mumps infection.
<b>Significant Result:</b>	Positive IgM indicates probable current or recent mumps infection.
<b>Limitations:</b>	(1) 30% of primary mumps may be sub-clinical. (2) Mumps infection can occur without parotitis. (3) Parotid swelling may have other viral/bacterial causes (Coxsackie, Echo, Parainfluenza, Influenza A, HSV, and <i>S. aureus</i> ). (4) Parotid pain or swelling may have a non-infectious cause.
<b>Turnaround Time:</b>	3 days
<b>Sample and Volume:</b>	Whole blood (10-15mL) collected aseptically in a red top vacutainer tube or single serum (2-3mL).
<b>Sample Collection:</b>	Serum collected 2-14 days post onset.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Due to the intense heat in the summertime, it is advisable to ship the specimens cold to prevent damage to the specimen. Whole blood samples may be sent on cool packs and not frozen. Serum samples, if not tested within 7 days or already frozen, should be shipped frozen to ASHL on dry ice. <u>If sent by courier:</u> <ol style="list-style-type: none"> <li>1. Blood and blood products sent in vacutainer tubes should first be placed in a plastic falcon tube.</li> <li>2. The specimen should then be placed in a plastic specimen bag with separate compartments for the submission form and the specimen.</li> <li>3. Pack the specimen and its form in absorbent material</li> <li>4. Mark the outer packaging “UN3373- Biological Substances, Category B.”</li> </ol> <u>If sent by mail:</u> <ol style="list-style-type: none"> <li>1. Blood and blood products sent in vacutainer tubes should first be placed in a plastic falcon tube.</li> <li>2. Check with the Post Office for current postage requirements.</li> <li>3. Wrap the submission form around the falcon tube, and place the falcon tube inside a Styrofoam container or cardboard mailer. Pack the specimen in absorbent material.</li> <li>4. Mark the outer packaging “UN3373- Biological Substances, Category B.”</li> </ol>

<b>Test Name:</b>	<b><u>Mumps, Molecular Typing PCR</u></b>
<b>Use of Test:</b>	Detection of mumps RNA virus
<b>Turnaround Time:</b>	2 days
<b>Sample:</b>	Nasopharyngeal swab, throat swab, CSF, buccal, sputum and urine (5mL).
<b>Sample Collection:</b>	<u>Nasopharyngeal swab:</u> The pharynx is swabbed vigorously with a cotton swab moistened with collection medium free of serum such as Hanks, and then placed in a transport container containing Hanks Buffered Saline Solution (HBSS). Specimens submitted should not be frozen. <u>Throat Swab:</u> The pharynx is swabbed vigorously with a cotton swab moistened with collection medium free of serum such as Hanks, and then placed in a transport container containing Hanks Buffered Saline Solution (HBSS). Samples submitted should not be frozen. <u>Urine:</u> Clean catch urine collected in a sterile container.
<b><u>Forms Required:</u></b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Use a triple packaging system for transporting by courier. Mark the outer packaging as “UN3373- Biological Substances, Category B.” <b><u>Ship cold</u></b>

**Neisseria meningitidis**

<b>Test Name:</b>	<b><u>Neisseria meningitidis Culture &amp; Serogrouping</u></b>
<b>Use of Test:</b>	Confirmation and serogrouping of <i>Neisseria meningitidis</i> .
<b>Limitations:</b>	Testing performed only on organisms isolated from normally sterile sites.
<b>Turnaround Time:</b>	5 days
<b>Sample:</b>	Pure culture on chocolate agar slant or culture plate.

**Specimen Collection:**

*Isolate*- In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure young culture on chocolate agar slant or culture plate. If the report is after hours, the county epidemiologist should also notify ADHS through the emergency after hours number.

**Sample Container:**

**Forms Required:**

It is recommended that the containers be insulated during very hot or very cold weather. The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

**Shipping Requirements:**

All cultures must be transported with minimum delay since viability is readily lost. Use a triple packaging system for transporting by courier or USPS. Mark the outer packaging “UN3373-Biological Substances, Category B.” Mark “DO NOT REFRIGERATE” on the outside of the package.

## **Norovirus**

**Test Name:**

**Norovirus, Molecular Typing PCR**

**Use of Test:**

Detection of norovirus RNA in stool or vomitus specimens

**Turnaround Time:**

7 days

**Sample:**

Vomit or stool. 10-15 mL in volume

**Sample Collection:**

Specimens should be collected as soon as possible after symptom onset, preferably within the first 24 hours of collection. Vomit 10-15mL must be submitted fresh in a sterile container. Stool 10-15mL can be submitted fresh, in a sterile container. **DO NOT FREEZE. DO NOT PUT IN PRESERVATIVES OR ANY OTHER MEDIA.**

**Forms Required:**

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

**Shipping Requirements:**

Use triple packaging system for transporting by courier only. Mark the outer packing "UN3373-Biological Substances, Category B". **Ship cold**

## **Rubella**

**Test Name:**

**Use of Test:**

**Test Includes:**

**Significant Result:**

**Limitations:**

**Turnaround Time:**

**Sample and Volume:**

**Sample Collection:**

**Forms Required:**

**Shipment Requirements:**

### **Rubella Antibody, IgM**

Determination of rubella infection

Rubella IgM EIA

Positive IgM indicates current or recent rubella infection.

IgM may be negative if the specimen is collected prior to the appearance of or before the third day after rash onset.

5 days

Whole blood (10-15mL) collected aseptically in a red top vacutainer tube or single serum (2-3mL).

Serum collected 3-7 days after appearance of rash.

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

Due to the intense heat in the summertime, it is advisable to ship the specimens cold to prevent damage to the specimen. Whole blood samples may be sent on cool packs and not frozen. Serum samples, if not tested within 7 days or already frozen, should be shipped frozen to ASHL on dry ice.

#### **If sent by courier:**

1. Blood and blood products sent in vacutainer tubes should first be placed in a plastic falcon tube.
2. The specimen should then be placed in a plastic specimen bag with separate compartments for the submission form and the specimen.
3. Pack the specimen and its form in absorbent material.
4. Mark the outer packaging "UN3373- Biological Substances, Category B."

#### **If sent by mail:**

1. Blood and blood products sent in vacutainer tubes should first be placed in a plastic falcon tube.
2. Check with the Post Office for current postage requirements.
3. Wrap the submission form around the falcon tube, and place the falcon tube inside a Styrofoam container or cardboard mailer. Pack the specimen in absorbent material.
4. Mark the outer packaging "UN3373- Biological Substances, Category B."

## Salmonella

**Test Name:**

**Use of Test:**

**Special Instructions:**

**Test Includes:**

**Turnaround Time:**

**Sample:**

**Forms Required:**

**Shipping Requirements:**

**Salmonella spp. Culture and Serotyping**

To screen for bacterial cause of diarrheal illness

Stool specimens must be properly submitted, with transport containers not overfilled and with transport medium not removed. Specimen jars must be tightly closed and not leaking when received. The time interval between collection of the specimen and receipt in the Lab must not be greater than 5 days.

Isolation and species identification of *Salmonella* species. Tests include serotyping for *Salmonella* spp. Problematic isolates are submitted to CDC for serotyping. Tests also include pulsed-field gel electrophoresis (PFGE).

14 days

*Isolate-* In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture on TSI or nutrient agar slant or culture plate

*Specimen-* If isolate not available, then collection of one of the following specimens is required for testing: Stool, Rectal swab in Cary-Blair media or blood.

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

Package and ship with label on outer packaging as “UN3373-Biological Substances, Category B”.

**Test Name:**

**Use of Test:**

**Special Instructions:**

**Test Includes:**

**Turnaround Time:**

**Sample and Volume:**

**Sample Container:**

**Sample Collection:**

**Forms Required:**

**Shipping Requirements:**

**Salmonella Isolation, Food**

To support epidemiologic evidence implicating a food as a possible source of illness.

Food samples must be submitted through local or state public health agencies as part of an outbreak investigation (one or more ill consumers). All food items must clearly be labeled with the name of the food source and date of collection

Isolation and identification of *Salmonella* species

3-7 days.

At least 200 grams of the solid product or 100 ml of liquid.

Original sample container as submitted by inspector or other sterile leak proof container.

Collect food aseptically and place in sterile whirlpack bags or other sterile, leak proof container. Keep all samples refrigerated except those samples received frozen which should be maintained in the frozen state.

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.

Triple package samples. Transport samples on ice or on prefrozen cold packs, in appropriate packaging.

## Shigella

<b>Test Name:</b>	<b><u>Shigella species Culture and Serotyping</u></b>
<b>Use of Test:</b>	To screen for bacterial cause of diarrheal illness.
<b>Special Instructions:</b>	Stool specimens must be properly submitted, with transport containers not overfilled and with transport medium not removed. Specimen jars must be tightly closed and not leaking when received. The time interval between collection of the specimen and receipt in the Lab must not be greater than 5 days.
<b>Test Includes:</b>	Isolation and species identification of <i>Shigella</i> spp. Tests include serotyping for <i>Shigella</i> spp. Problematic isolates are submitted to CDC for serotyping. Tests also include Pulsed-field gel electrophoresis (PFGE)
<b>Turnaround Time:</b>	5 days
<b>Sample:</b>	<i>Isolate</i> - In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture on TSI or nutrient agar slant or culture plate <i>Specimen</i> - If isolate not available, then collection of one of the following specimens is required for testing: Stool, Rectal swab in Cary-Blair media or blood.
<b>Forms Required:</b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Package and ship with label on outer packaging as “UN3373-Biological Substances, Category B”.

<b>Test Name:</b>	<b><u>Shigella Isolation, Food</u></b>
<b>Use of Test:</b>	To support epidemiologic evidence implicating a food as a possible source of illness.
<b>Special Instructions:</b>	Food samples must be submitted through local or state public health agencies as part of an outbreak investigation (one or more ill consumers). All food items must clearly be labeled with the name of the food source and date of collection
<b>Test Includes:</b>	Isolation and identification of <i>Shigella</i> species.
<b>Turnaround Time:</b>	3-7 days.
<b>Sample and Volume:</b>	At least 200 grams of the solid product or 100 ml of liquid.
<b>Sample Container:</b>	Original sample container as submitted by inspector or other sterile leak proof container.
<b>Sample Collection:</b>	Collect food aseptically and place in sterile whirlpack bags or other sterile, leak proof container. Keep all samples refrigerated except those samples received frozen which should be maintained in the frozen state.
<b>Forms Required:</b>	The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type.
<b>Shipping Requirements:</b>	Triple package samples. Transport samples on ice or on prefrozen cold packs, in appropriate packaging.

## Vibrio

**Test Name:**

**Use of Test:**

**Special Instructions:**

**Turnaround Time:**

**Sample and Volume:**

**Sample Collection:**

**Forms Required:**

**Shipping Requirements:**

**Vibrio spp. Culture**

To screen for bacterial cause of diarrheal illness.

Cultures suspected to contain *Vibrio cholera* are tested with commercial biochemical systems. Cultures presumptively identified as *Vibrio cholera* will be tested against specific antisera to determine the serogrouping of the isolate. *Vibrio cholera* strains will fall into two groups based on this testing. Serogroup O1 is associated with epidemic cholera; non-O1 strains may cause cholera-like illness, but are not involved in epidemics. Serogroup O1 isolates are typed at ASHL and sent to CDC for confirmation.

5 days. If sent to CDC for Serogroup O1 confirmation turnaround time may vary  
Stool specimen (approximately one gram)

*Isolate*- In the event of an outbreak, the hospital or commercial laboratory, should forward the isolate to ASHL. The isolate should be a pure culture on TSI or nutrient agar slant or culture plate

*Specimens*- Stool specimens should be collected early, preferably within 24 hours of onset of illness, and before the administration of antibiotics. Rectal swabs or fecal material should be placed in Cary-Blair transport medium. The Arizona State Public Health Laboratory will provide agencies with Cary-Blair Medium.

The ASHL Microbiology Submission Form must be submitted with each specimen. The fields that are required for specimen processing include: Patient first and last name, age, sex, date of birth, submitting agency name, specimen collection date, and specimen type. Specimens held in Cary-Blair Medium should be transported at 5-25°C. DO NOT REFRIGERATE. Use triple packaging system for courier or USPS. Mark outer packaging as “UN3373-Biological Substances, Category B”.

## **Bacterial Typing**

**Test Name:**

**Bacterial Typing, Pulsed Field Gel Electrophoresis (PFGE)**

**Use of Test:**

To determine if isolates from different patients or sources (i.e. patient and environmental isolates) have a common origin. Test is very discriminatory, and is primarily used in food related outbreaks.

ASHL bacteriology laboratory can currently perform PFGE on all confirmed isolates of *Salmonella* spp., *Shigella* spp., *E.coli* O157:H7, *Campylobacter* spp. and *Listeria* spp.

AHSL routinely performs PFGE on all isolates if these organisms have been identified.

PFGE for all other organisms isolated would be sent to CDC for PFGE.

In the event of a Nosocomial outbreak, and by request only, ASHL can conduct PFGE on *Acinetobacter* spp., *Pseudomonas* spp., *Burkholderia* spp.

PFGE is not a diagnostic test and is used in conjunction with epidemiological findings that result from intense investigation.

Accurate identification of all isolates must be confirmed prior to PFGE testing.

Results are interpreted based on banding patterns

**Test Includes:**

Bacterial strain typing using restriction endonucleases (enzyme) digestion of bacterial chromosomal DNA.

**Turnaround Time:**

1 week for pure cultures. Turn around time is delayed if patient specimens are submitted or if the isolate submitted is contaminated.

**Sample:**

Pure isolates must be received on agar slants

## **SECTION V: ARIZONA STATE PUBLIC HEALTH LABORATORY MISCELLANEOUS DOCUMENTS**

### **SPECIMEN REJECTION POLICY**

The State Laboratory currently has the following policy for rejection of laboratory specimens and/or requested examinations. The State Laboratory will usually NOT examine clinical/reference specimens if the following circumstances exist:

- The quantity of the specimen is not sufficient for examination
- The specimen was too long in transit between the time of collection and receipt in the laboratory
- The specimen was broken or leaked in transit
- Clinical/epidemiological information submitted with the specimen was either insufficient or incomplete
- Specimen was submitted in an improper container, transport media or preservative
- Blood specimens are hemolyzed or contaminated
- The identifier on the specimen does not match the identifier on the submission form, or there is no identification on the specimen
- Materials for rabies examination are too decomposed to test
- Test is available at a hospital/independent laboratory and has been discontinued by the State Laboratory
- Reference cultures are contaminated

Exceptions to this policy will be considered due to extenuating circumstances; however, final approval to make an exception will only be made by the Laboratory Director, Bureau Chief or Assistant Bureau Chief delegated this responsibility.

# ASHL MICROBIOLOGY SUBMISSION FORM

For an editable version of the form, go to <http://www.azdhs.gov/lab/micro/pdf/micro-submission-form.pdf>



## Bureau of Laboratory Services

250 N. 17<sup>th</sup> Avenue Phoenix, Arizona 85007-3231  
 Tel: (602) 542-1188 Fax: (602) 364-0758  
 Victor Waddell, Ph.D., Bureau Chief

For Department Use Only

### PATIENT INFORMATION

Last name: \_\_\_\_\_ First name: \_\_\_\_\_ MI: \_\_\_\_\_  
 DOB (MM/DD/YYYY): \_\_\_\_\_ Age: \_\_\_\_\_ Sex:  M  F  T Patient ID: \_\_\_\_\_  
 Street address: \_\_\_\_\_ City: \_\_\_\_\_ State: \_\_\_\_\_ Zip: \_\_\_\_\_ County: \_\_\_\_\_  
 Ethnicity: Hispanic  Yes  No Race:  White  African American  Asian  American Indian/Alaska Native  Other

### SUBMITTING AGENCY INFORMATION

Agency name: \_\_\_\_\_ Agency ID Code: \_\_\_\_\_  
 Street address: \_\_\_\_\_ City: \_\_\_\_\_ State: \_\_\_\_\_ Zip: \_\_\_\_\_ County: \_\_\_\_\_  
 Contact name: \_\_\_\_\_ Tel: \_\_\_\_\_

### ORDERING PROVIDER INFORMATION

Provider name: \_\_\_\_\_ Tel: \_\_\_\_\_  
 Facility name: \_\_\_\_\_ Tel: \_\_\_\_\_  
 Street address: \_\_\_\_\_ City: \_\_\_\_\_ State: \_\_\_\_\_ Zip: \_\_\_\_\_ County: \_\_\_\_\_

### SPECIMEN INFORMATION & TYPE

Collection date: \_\_\_\_\_

<input type="checkbox"/> Serum <input type="checkbox"/> Acute <input type="checkbox"/> Convalescent	<input type="checkbox"/> Plasma	<input type="checkbox"/> Whole blood	<input type="checkbox"/> CSF
<input type="checkbox"/> Swab, site: _____	<input type="checkbox"/> Urine	<input type="checkbox"/> Sputum	<input type="checkbox"/> Stool
Other, specify: _____	<input type="checkbox"/> Tissue, specify: _____	<input type="checkbox"/> Wound, site: _____	<input type="checkbox"/> Body fluid, specify: _____

Clinical  Reference:  Broth  Isolate Reason for testing:  Outbreak  Surveillance  Post Mortem  Screening  Diagnostics  
 Outbreak name: \_\_\_\_\_

<p><b>VIROLOGY</b></p> <p><input type="checkbox"/> CMV Culture  <input type="checkbox"/> Enterovirus Culture  <input type="checkbox"/> Herpes Culture  <input type="checkbox"/> Influenza  <input type="checkbox"/> Norovirus PCR  <input type="checkbox"/> Reference Virus Culture  <input type="checkbox"/> Respiratory Virus Culture  <input type="checkbox"/> Other _____</p> <p><b>SELECT AGENTS**</b></p> <p><input type="checkbox"/> Avian Influenza H5N1  <input type="checkbox"/> <i>Bacillus anthracis</i>  <input type="checkbox"/> <i>Brucella</i> spp.  <input type="checkbox"/> <i>Burkholderia</i> spp.  <input type="checkbox"/> <i>Francisella tularensis</i>  <input type="checkbox"/> <i>Orthopox</i>  <input type="checkbox"/> <i>Q</i> Fever  <input type="checkbox"/> SARS  <input type="checkbox"/> <i>Yersinia pestis</i></p> <p><small>**Please refer to the Guide to Laboratory Services: Microbiology, Section 8 for the definition of select agents and the testing available at Arizona State Laboratory</small></p>	<p><b>BACTERIOLOGY</b></p> <p><input type="checkbox"/> <i>Bordetella pertussis</i>  <input type="checkbox"/> <i>Campylobacter</i> spp.  <input type="checkbox"/> <i>Clostridium botulinum</i> toxin  <input type="checkbox"/> <i>Corynebacterium diphtheriae</i>  <input type="checkbox"/> Enteric culture  <input type="checkbox"/> <i>Escherichia coli</i> / Shigatoxin  <input type="checkbox"/> <i>Haemophilus influenzae</i>  <input type="checkbox"/> <i>Legionella</i> spp.  <input type="checkbox"/> <i>Leptospira</i> spp.  <input type="checkbox"/> <i>Listeria</i> spp.  <input type="checkbox"/> <i>Neisseria gonorrhoeae</i>  <input type="checkbox"/> <i>Neisseria meningitidis</i>  <input type="checkbox"/> <i>Salmonella</i> spp.  <input type="checkbox"/> <i>Shigella</i> spp.  <input type="checkbox"/> <i>Streptococcus pneumoniae</i>  <input type="checkbox"/> <i>Vibrio</i>  <input type="checkbox"/> VISA/VRSA  <input type="checkbox"/> <i>Yersinia</i> (Non-pestis) Culture  <input type="checkbox"/> Other: _____</p> <p><b>PARASITOLOGY†</b></p> <p><input type="checkbox"/> Arthropod ID  <input type="checkbox"/> Blood/Tissue  <input type="checkbox"/> Intestinal  <input type="checkbox"/> Pinworm  <input type="checkbox"/> Worm ID</p> <p><small>† For malaria testing please collect patient travel history</small></p>	<p><b>SEROLOGY</b></p> <p><input type="checkbox"/> <i>Borrelia burgdorferi</i> EIA (Lyme)  <input type="checkbox"/> <i>Brucella</i> Tube Ag.  <input type="checkbox"/> Coccidioides Serology Panel <input type="checkbox"/> IDTP <input type="checkbox"/> IDCF  <input type="checkbox"/> Dengue IgM EIA  <input type="checkbox"/> Diagnostic Hepatitis Panel EIA <input type="checkbox"/> HBsAG <input type="checkbox"/> HBcIgM <input type="checkbox"/> HAV IgM  <input type="checkbox"/> <i>Francisella tularensis</i> Tube Ag.  <input type="checkbox"/> <i>Hantavirus</i> IgG EIA  <input type="checkbox"/> <i>Hantavirus</i> IgM EIA  <input type="checkbox"/> Hepatitis Anti-HAV IgM  <input type="checkbox"/> Hepatitis Anti-Core IgM  <input type="checkbox"/> Hepatitis Anti-HCV  <input type="checkbox"/> Hepatitis HBsAG  <input type="checkbox"/> <i>Measles</i> IgM EIA  <input type="checkbox"/> <i>Mumps</i> IgM EIA  <input type="checkbox"/> Rickettsial Panel IFA <input type="checkbox"/> Rickettsial <i>Q</i> Fever <input type="checkbox"/> Rickettsial Spotted Fever  <input type="checkbox"/> Rickettsial Typhus Fever</p> <p><input type="checkbox"/> <i>Rubella</i> IgM EIA  <input type="checkbox"/> St. Louis Encephalitis EIA  <input type="checkbox"/> <i>Treponema pallidum</i> CSF VDRL  <input type="checkbox"/> <i>Treponema pallidum</i> Serum TP-PA, RPR  <input type="checkbox"/> Western Equine Encephalitis EIA  <input type="checkbox"/> West Nile Virus EIA  <input type="checkbox"/> <i>Yersinia pestis</i> PHA  <input type="checkbox"/> Other: _____</p>
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Submitting Lab Findings or Preliminary ID: \_\_\_\_\_  
*\*Prior notification is required for: Bacillus anthracis, Bordetella pertussis, Brucella spp., Clostridium botulinum toxin, Corynebacterium diphtheriae, emerging or exotic diseases, Francisella tularensis, Hantavirus, Borrelia burgdorferi (Lyme), Measles, Mycobacteria NAA, Rubella, or Yersinia pestis testing. CALL: (602) 364-3676*

ALL FIELDS HIGHLIGHTED IN YELLOW ARE REQUIRED FOR SPECIMEN PROCESSING. IN ADDITION, AT LEAST ONE TEST MUST BE REQUESTED.

Patient address and telephone number are required, when available, per R9-6-204(B3) <http://www.azdhs.gov/lab/micro/index.htm>

Updated 10/20/2010

## REQUESTING COLLECTION KITS AND MAILING CONTAINERS

Supplies ordered from the Arizona State Laboratory are to be used **ONLY** to submit specimens to the State Laboratory. There are two Request for Materials forms currently in use: a Newborn Screening Supplies Request Form and a Request Form for all other supplies available from the State Laboratory. Supplies can be requested by mailing, faxing, calling, or emailing the Receiving Section at:

Arizona Department of Health Services  
Bureau of State Laboratory Services  
ATTN: Receiving Section  
250 North 17th Avenue  
Phoenix, AZ 85007  
Fax (602) 364-0758  
Phone (602) 542-1190  
Email [labreceiving@azdhs.gov](mailto:labreceiving@azdhs.gov)

Please request materials before they are required as the expected turn around time per order is FIVE business days. Most materials do have a limited shelf life; therefore, only order what will be used before the expiration date. Please do not use expired kits or any kits in which the medium has changed characteristics. Dispose of the media properly and order replacement supplies. The following table provides information regarding submission forms, kit contents and expiration period of each kit. Submitters may use the Request for Materials Form to order entire kits, as well as individual components.

KIT	CONTENTS	SHELF LIFE
Enteric Kit	Instruction Sheet Baggie Metal Container Cardboard Mailer Media: Cary Blair Store +20 to +25°C	6 months
Influenza Kit	Microbiology Submission Form Instruction Sheet N/P Swab Media: Hanks Blue Top. Store +2 to +8°C	2 months
Leptospira Culture Media	Leptospira Media Instructions	6-12 months
Ova & Parasite Kit	Instruction Sheet Baggie Metal Container Cardboard Mailer Media: PVA & Formalin. Store +20 to +30°C	1-2 years
Pertussis Kit	Microbiology Submission Form Instruction Sheet Polyester N/P Swab (metal handle) Media: Regan Lowe. Store +2 to +8°C	2 months
Tuberculosis Kit	Sputum Vial Metal Container Cardboard Mailer Store +20 to +25°C	